Synthesis of the Securinega Alkaloids (±)-Norsecurinine and (±)-Nirurine from 3-Hydroxypyridine.

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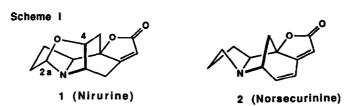
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Dedicated to Derek H. R. Barton on the occasion of his 75th birthday

Abstract: Treatment of the dihydropyridine 5 with fluoride anion resulted in in situ formation of the allenyldiene 4 which cyclized to the azabicyclo[2.2.2]octene 9. Subsequent elaboration to the alcohol 24 and rearrangement gave norsecurinine 2. The intermediate diol 16 was converted into prenirurine 34 which upon conversion into an N-oxide rapidly rearranged to give traces of nirurine 1 and norphyllanthine 38.

(+)-Nirurine 1 was isolated from *Phyllanthus niruri* L., and its pentacyclic structure elucidated by X-ray crystallography. Nirurine 1 is biogenetically related to norsecurinine 2 (also isolated from *Phyllanthus*). Norsecurinine 2 has been synthesized, however there are no reported synthetic studies on nirurine 1, nor any structural relationships established between the azabicyclo[2.2.2]octane core of nirurine and the azabicyclo[3.2.1]octane core of norsecurinine.



The strategy we have developed to construct the azabicyclo[2.2.2]octane (isoquinuclidine) core of nirurine depends upon the generation of the diene 4/4a, and its stereospecific intramolecular trapping by an allenyl ester to produce the core skeleton and the fused butenolide 3 in a single step, Scheme II.3 In turn, we envisioned that the allenyl ester could be generated in situ from the propargylic ester 5 by treatment with fluoride anion. The stereochemical outcome of the intramolecular 2+4 cycloaddition to give 3, requires that the allyl side chain occupy the face opposite that of the allenyl substituent. This appears to be the more favorable situation (steric hindrance), and it was anticipated that 4, rather than 4a, would determine the correct stereochemistry required for the eventual pyrrolidine ring. The 1,2-dihydro-2-allylpyridine 5 should be readily available from 6, which is the simple propargylation product of the 3-hydroxypyridine 7. While we initially

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examined the successful prototype situation where R = H, leading to 3 (R = H), there remains the somewhat difficult problem of selectively introducing the required oxygen functionality in a regiospecific manner on the bridging C=C double bond, where both ends have inductively electron withdrawing substituents. The only difference between the ends of the double bond is the degree of steric hindrance.

3-Hydroxy-5-methoxypyridine 7 (R = OMe) was converted into the ester 6 (R = OMe, $X = SiMe_3$), and treated with methyl chloroformate/allyl tri-n-butyl stannane to give the adduct 5 as a mixture containing other allyl regioisomers. Treatment of the mixture with fluoride anion did not give 3 (R = OMe). The only identifiable product was the starting pyridine 7. Consequently, we were restricted to the prototype series (R = H).

4-Trimethylsilyl-2-butynoic acid 8 was prepared by deprotonation of 2butynoic acid with n-BuLi/TMEDA/-78 °C, followed by the addition of trimethylsilyl chloride. The acid 8 is rather sensitive, and is best stored in a freezer and used 3-Hydroxypyridine was treated with 4-trimethylsilyl-2-butynoic within a few days. acid DCC/CH₂Cl₂ to give the labile ester 6 (78%). Immediate exposure of 6 to the Yamaguchi allylation conditions⁴ (ClCO₂Me/allyltri-n-butylstannane/0-25 °C gave the 2-allyl-1,2-dihydropyridine adduct 5 (81%). No other regioisomers could be detected (1H NMR). Fluoride ion induced desilylation of 5 (KF. 2H2O/MeOH/AcOH) gave the azabicyclo[2.2.2]octane 9 (45%) as a single stereoisomer, presumably via the intermediate allenyl-diene 4. We could not detect any stereoisomer that would arise from the endo-allyl intermediate 4a. The only significant by-product was protodesilylation of 5 (replacement of SiMe₃ by H). If an anhydrous fluoride source was used (CsF or n-Bu₄N+F-) none of the required product 9 could be isolated, only extensive decomposition was observed. Consequently, we were restricted to the protic system with the limitation of competing protodesilylation. The complete sequence of transformations from 6 to 9 can be carried out without purification of the intermediates, Scheme III.

Selective hydroboration (1° vs 2°) of the vinyl functionality in 9 was readily achieved by exposure of 9 to dissoamylborane/THF/-23 °C followed by oxidative work-up to give the alcohol 10 (89%). The allylically disubstituted double bond showed no inclination towards competitive hydroboration. Attempted deprotection of the carbamate 10 using trimethylsilyliodide resulted in decomposition. Whereas, conversion of 10 into its derived p-toluenesulfonyl ester 11, followed by treatment with hydrogen bromide in acetic acid containing a bromine scavenger (cyclohexene) gave the core structure 3 (77%).

Our initial plan (which failed) is worth mentioning if only for the fact that it reemphasizes the inertness of the allylically disubstituted double bond in 3 (and similar structures) towards electrophilic addition chemistry. Oxidation of 10 using the Swern-Moffatt procedure gave the aldehyde 12. Treatment of the aldehyde with hydrogen bromide in acetic acid gave the unstable carbanolamine 13 as an anomeric mixture (¹H NMR). The derived methyl ether is a single epimer. All attempts [HgO/Br₂/CCl₄, PhSeCl, PhSeBr, NBS, NIS, Br₂, Hg(OCOCF₃)₂, KI/I₂, I₂/AgOAc/AcOH, MCPBA etc, and photolysis] failed to induce direct cyclization of 13 into nirurine 1. The only product that could be isolated from these cyclization attempts was the amide 14 (40%, from the NBS reaction). Thus, this direct and simple route, could not be realized. Scheme IV.

In the course of this part of the research it was found that the only addition reaction to the unconjugated C=C double bond in 10 could be achieved by osmylation using OsO₄(cat)/NMNO/t-BuOH/THF-water⁵ to give a cis-diol (at this stage of unspecified stereochemistry). The cis-diol was converted into its acetonide derivative and oxidized to give the crystalline aldehyde 15 (X-ray). This structure

confirmed the stereochemistry of the osmylation (approach from the trigonal butenolide side), and the stereochemistry of the aldehyde side chain (allyl, 4 into 9, Scheme III).

As emphasized above, the disubstituted double bond in 3 proved to be extremely reluctant to undergo electrophilic addition, presumably because of the strongly inductively electron-withdrawing allylic-N and O-substituents. The only useful functionalization was achieved by treatment of 3 with OsO₄(cat)/NMNO/ acetone-water to give the cis-diol 16 (90%). Unfortunately, this is the incorrect configuration at C-4, and consequently we were faced with the difficult task of inverting at C-4 in a molecule where SN2 chemistry is obviously sterically encumbered, combined with the problem of differentiating between the two secondary hydroxyl groups.

After considerable experimentation it was found that pivaloylation of the diol 16 gave a mixture of monopivaloates 17 and 18 (1:2) (100%). If a mixture of 17 and 18 is allowed to stand in methanol for a few minutes the ¹H NMR spectrum indicated that rapid equilibration takes place to give predominantly 18. Swern-Moffatt oxidation of the mixture of 17 and 18 gave 19, along with a small amount of the isomer 20. Evidently 18 is more rapidly oxidized than 17. Consequently while pivaloylation of 16 is not regiospecific, the subsequent equilibration allows the ketone 19 to be made without any separation from isomeric compounds. Reduction of 19 gave the inverted alcohol 21 (64% overall from 17/18). Deoxygenation of 21 using the Barton procedure via the pentafluorophenolthiono ester derivative 22 (100%) and subsequent treatment with n-Bu₃SnH/AIBN gave 23 (100%). Treatment of the ester 23 with NaOMe/MeOH gave the alcohol 24 (94%). The alcohol 24 (94%) cleanly rearranged to norsecurinine 2 (91%, overall yield of 10.5% through 13 steps from 3-hydroxypyridine) on exposure to standard mesylation conditions, Scheme V.

a) OsO₄ (cat)/N-methylmorpholine-N-oxide/acetone, water (90%). b) Bu^tCOCl/DMAP, Et₃N/CH₂Cl₂ (100%). c) DMSO/(ClCO)₂/Et₃N. d) NaBH₄/MeOH (64% from 16). e) C₆F₅OCSCl/DMAP/ CH₂Cl₂ (100%). f) n-Bu₃SnH/AIBN/PhH reflux (100%). g) NaOMe/MeOH (94%). h) MeSO₂Cl/CH₂Cl₂/Et₃N (91%).

The rearrangement of the seco-nirurine skeleton 24 into the norsecurinine skeleton 2 may have possible biogenetic relevance since both alkaloids occur in the same plant. This same type of rearrangement can be conducted on the diol 16. Treatment of 16 with the Mitsunobu reagent PPh3/EtO2CN=NCO2Et/THF did not give the desired inverted alcohol, but instead the rearranged 8-hydroxynorsecurinine 26 (87%) was formed (structure by X-ray). We considered the possibility that the derived N-oxide 27 would undergo rearrangement into the O-alkylhydroxylamine 28, which could be oxidized to 29, and eliminated to the nitrone 30.1 The nitrone 30 should cyclize to 31, and undergo conjugate addition to give the N-oxide of hydroxynirurine 32. Treatment of 8-hydroxynorsecurinine 26 with m-chloroperoxy benzoic acid gave the N-oxide 27 (100%), which cleanly rearranged to the required O-alkyl hydroxylamine 28 (99%) when heated in xylene at reflux. When 28 was treated with m-chloroperoxybenzoic acid an unstable polar material was formed, which upon heating gave a complex mixture that did not contain (1H NMR, carbanol-amine CH) any of the rearranged product 32, Scheme VI.

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Scheme VI

a) Mitsunobu conditions (87%). b) m-ClC₆H₄CO₃H/MeOH (100%). c) Xylene reflux (99%).

Oxidation of 24 using the Swern-Moffatt conditions gave the ketone 33, which was directly reduced with sodium borohydride to give the inverted alcohol 34 (82%, from 24). Treatment of 34 with a variety of oxidizing agents [NBS, NIS, Hg(OCOCF₃)₂, Hg(OAc)₂, Pb(OAc)₄ etc] either destroyed 34 to give numerous products none of which were 1 or 34, or was recovered unchanged. It was found that treatment of 34 with m-chloroperoxybenzoic acid in methanol gave the unstable N-oxide 35, which rapidly rearranged to 38, presumably via the Cope elimination product 36/37. The N-oxide 35 is more stable in dichloromethane, and treatment with trifluoroacetic anhydride gave small amounts of nirurine 1 (ca.10%), but largely 38. In view of the low yield of 1, because of the competing rearrangement, it seems likely that 34 is not the biogenetic precursor to 1, and that aminal formation (oxidation adjacent to nitrogen) takes place at an earlier stage.

Summary

The simple strategy depicted in Scheme II provides a short route to azabicyclo[2.2.2]octane skeleton 3. While the subsequent elaboration of 3 to give norsecurinine 2 was straightforward, the conversion of 3 into nirurine was extremely inefficient because of the difficulty experienced during attempts to oxidize prenirurine 34 to the iminium ion 39. This perhaps illustrates the paucity of methods that are available that directly dehydrogenate amines to iminium species in a controlled manner. We are currently developing a new method based upon hypervalent iodine chemistry to address this specific problem.

a) DMSO/(ClCO)2/Et3N. b) NaBH4/MeOH (82% from 24), c) m-ClC6H4CO3H/CH2Cl2.

Experimental Section

4-Trimethylsilyl-2-Butynoic acid 8. A stirred solution of tetramethyl ethylenediamine (4.82 ml, 31.9 mmol) in dry pentane (100ml) under argon was treated with a 2.5 M solution of n-BuLi in hexanes (18.4 ml, 46.0 mmol) at 25 °C. After 1 h, the reaction mixture was cooled to -78 °C, and a solution of 2-butynoic acid (1.75 g, 20.8 mmol) in THF (25 ml) was added via cannula. After 2 h, the resulting yellow suspension was treated with trimethylsilyl chloride (10.6 ml, 85.4 mmol). After a further 1.5 h at -78 °C, the mixture was allowed to warm to room temperature. The reaction mixture was poured into 10% aqueous HCl (90 ml), and extracted with Et₂O (2x50 ml). The combined organic layers were dried (MgSO₄), and concentrated in vacuo to give a yellow oil. Kugelrohr distillation provided the acid 8 (3.175 g, 98%) as a colorless oil, which solidified on storage in the freezer, B.p. 123 °C/0.45 mm Hg. IR (film) 3000 (br), 2951, 2219, 1678, 1408, 1286, 1251, 844 cm⁻¹. ¹H NMR (300 MHz, C₆D₆) & 9.4 (1H, br s), 1.12 (2H, s), -0.14 (9H, s). ¹³C NMR (75 MHz, C₆D₆) & 158.66, 92.19, 73.09, 7.38, -2.34. CIMS, m/e 157 (MH+, base), 139, 113. HRMS, m/e calcd. for C₇H₁₃O₂Si 157.0685. Found 157.0704.

Azabicyclo[2.2.2]octane Diels-Alder adduct 9. A stirred suspension of 3-hydroxypyridine (0.717 g, 7.54 mmol) in dry CH₂Cl₂ (45 ml) under argon was treated with a solution of dicyclohexylcarbodiimide (1.556 g, 7.54 mmol) in dry CH₂Cl₂ (20 ml), followed by a solution of 4-trimethylsilyl-2-butynoic acid 8 (1.178 g,

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7.54 mmol) in dry CH₂Cl₂ (20 ml). After 10 min, the mixture was treated with saturated aqueous NaHCO₃, and diluted with CH₂Cl₂ (20 ml). The organic layer was separated, dried (Na₂SO₄), and concentrated *in vacuo* to give a brown oil which was rapidly purified by silica gel chromatography (10:1 dichloromethane: ethyl acetate) to provide the ester 6 as a yellow oil (1.373 g, 78%). It was used immediately in the next reaction to avoid decomposition.

The ester 6 from the above sequence was dissolved in dry CH₂Cl₂ (20 ml), cooled to 0 °C, and treated with methyl chloroformate (0.546 ml, 7.07 mmol), followed by allyltri-n-butylstannane (1.82 ml, 5.46 mmol). The resulting yellow solution was allowed to warm to 25 °C, and was stirred under argon for 3 h. The mixture was concentrated in vacuo, and the residue purified by silica gel chromatography eluting with hexane followed by dichloromethane to provide the carbamate 5, (1.478 g, 81%), as a yellow oil. It was used immediately in the next sequence to avoid decomposition.

The carbamate 5 was dissolved, with stirring, in methanol (300 ml), and treated with potassium fluoride dihydrate (4.18 g, 44.40 mmol), followed by acetic acid (0.266 ml, 4.44 mmol). After 17 h, the mixture was concentrated in vacuo, and the residue partitioned between CH₂Cl₂ (20 ml) and water (20 ml). The aqueous layer was extracted with CH₂Cl₂ (2x20 ml), and the combined organic layers were dried (Na₂SO₄) and concentrated in vacuo to give a brown oil, which was purified by silica gel chromatography to give the desired adduct 9 (0.517 g, 45%) as a pale yellow oil. IR (CH₂Cl₂) 2920, 1760, 1695, 1659 cm⁻¹. ¹H NMR (CD₃SOCD₃, 500 MHz at 70 °C) δ 6.61 (1H, dd, J = 8.4, 6.4 Hz), 6.47 (1H, d, J = 8.4 Hz), 5.97 (1H, t, J = 1.9 Hz), 5.82 (1H, m), 4.95-5.03 (3H, m), 3.63 (3H, s), 3.48 (1H, t, J = 6.0 Hz), 2.92 (1H, dt, J = 18.6, 1.9 Hz), 2.32-2.53 (3H, m). ¹³C NMR (125 MHz, C₆D₆ at 70 °C) δ 172.07 (s), 168.94 (s), 155.19 (s), 135.29 (d), 131.41 (d), 131.18 (d), 116.44 (t), 112.25 (d), 86.89 (s), 61.51 (d), 52.31 (q), 49.07 (d), 37.21 (t), 31.55 (t). CIMS, m/e 262 (MH+), 220 (base), 192, 154, 128, 96. HRMS, m/e calcd for C₁₄H₁₅NO₄ 262.1080. Found 262.1068.

Hydroboration of 9. A solution of BH₃,THF (1.0 M in THF, 17.8 ml, 17.8 mmol) at 0° C under argon was treated 2-methyl-2-butene (2.0 M in THF, 17.8 ml, 35.6 mmol) and the resulting solution left at 0 °C for 1 h. This solution was added via cannula to a stirred solution of the alkene 9 (0.930 g, 3.56 mmol) in dry THF (50 ml) at -23 °C under argon. After 2.5 h, the reaction mixture was allowed to warm to room temperature, and 30% aqueous H₂O₂ (15 ml), and 2 M aqueous NaOH (15 ml) were carefully added. After a further 0.5 h, the reaction mixture was treated with 2M aqueous Na₂SO₃. When effervescence had ceased, the mixture was extracted with EtOAc (3x20 ml), dried (Na₂SO₄), and concentrated in vacuo to give a colorless oil. Purification by silica gel chromatography, eluting with ethyl acetate gave the alcohol 10 as a colorless oil (0.893 g, 89%). IR (CH₂Cl₂) 3620, 3500, 2930, 1770, 1690, 1670 cm $^{-1}$. ¹H NMR (500 MHz, CD₃SOCD₃ at 80 °C) δ 6.58 (1H, dd, J = 8.4 and 6.3 Hz), 6.45 (1H, d, J = 8.4 Hz), 5.93 (1H, s), 4.98-5.06 (1H, m), 4.05 (1H, br s), 3.60 (3H, s), 3.50-3.32 (3H, m), 2.88 (1H, d, J = 18.9 Hz), 2.47 (1H, d, J = 18.9 Hz), 1.70-1.40(4H, m). ¹³C NMR (75 MHz, CD₃SOCD₃ at 80 °C) δ 172.08 (s), 170.29 (s), 154.54 (s), 131.53 (d), 129.94 (d), 110.83 (d), 86.71 (s), 60.53 (t), 60.36 (d), 51.95 (q), 48.28 (d), 31.24 (t), 29.16 (t), 28.69 (t). CIMS m/e 280 (MH+), 262, 220, 192, 154, 134, 106 (base).

HRMS, m/e calcd for C₁₄H₁₈NO₅ 280.1185. Found 280.1192.

Desoxadehydronirurine 3. A stirred solution of the alcohol 10 (1.97 g, 7.06 mmol) in dry CH_2Cl_2 (107 ml) was treated with triethylamine (1.08 ml, 7.77 mmol), followed by 4-dimethylaminopyridine (947 mg, 7.77 mmol) and p-toluenesulphonyl chloride (1.48 g, 7.77 mmol). After 5 h, the reaction mixture was diluted with CH_2Cl_2 (100 ml) and washed with 5% aqueous HCl, and water. The organic layer was dried (MgSO₄), and concentrated in vacuo to give the crude tosylate 11 as colorless foam. The crude tosylate was used directly in the next stage. Rf = 0.65 (silica gel, EtOAc).

The crude tosylate 11 from the above sequence was suspended in cyclohexene (26 ml) and treated with 30% HBr in acetic acid (107 ml) under argon. After 17 h, the mixture was added slowly to an excess of saturated aqueous Na_2CO_3 (50 ml), and the resulting cloudy suspension extracted with dichloromethane (5x20 ml). The combined organic layers were dried (MgSO₄), and concentrated in vacuo to give a pale yellow oil, which was purified by silica gel chromatography (EtOAc/MeOH 10:1, then 7:3) to give the amine 3 as a colorless solid (1.10 g, 77% overall). M.p. 57-59 °C. IR (film) 2965, and 1760 cm⁻¹. ¹H NMR (300 MHz, CDCl₃) δ 6.50 (1H, dd, J = 8.5, 5.1 Hz), 6.31 (1H, d, J = 8.5 Hz), 5.70 (1H, t, J = 1.8 Hz), 3.95 (1H, dd, J = 4.6, 1.6 Hz), 3.14 (2H, m), 3.00-2.93 (1H, dt, J = 18.4, 1.9 Hz), 2.62-2.53 (1H, m), 2.46-2.38 (1H, dt, J = 18.4, 2.0 Hz), 1.94-1.90 (1H, m), 1.77-1.54 (3H, m). ¹³C NMR (75 MHz, CDCl₃) δ 174.26 (s), 171.35 (s), 135.14 (d), 128.22 (d), 109.72 (d), 88.46 (s), 66.23 (d), 54.30 (d), 53.83 (t), 30.73 (t), 27.93 (t), 24.81 (t). EIMS, m/e 203 (M+), 134, 106, 78, 70 (base). HRMS calcd. for C₁₂H₁₃NO₂ 203.0939, found 203.0946. Anal calcd.for C₁₂H₁₃NO₂. C, 70.91; H, 6.45; N, 6.89. Found C, 70.78; H, 6.34; N, 6.85%.

Dihydroxynirurine 16. A stirred solution of the alkene 3 (0.040 g, 0.196 mmol) in acetone/water (5:2) (2 ml) was treated with 4-methylmorpholine-N-oxide [0.058 g, 0.490 mmol, followed by osmium tetroxide (2.5 wt % in t-BuOH) (0.040 ml, 0.0039 mmol]. After 4 h, the reaction mixture was treated with saturated aqueous Na₂SO₃, and stirred at room temperature for a further 0.5 h. The reaction mixture was diluted with water and extracted with n-BuOH (3x20 ml), dried, (Na2SO4), and evaporated to give an off white solid, which was purified by silica gel chromatography, eluting with CH₂Cl₂/MeOH (10:1) to give the diol 16 (0.042 g, 90%) as a colorless oil, which crystallized on standing. M.p 152-162 °C (dec). IR (KBr) 3600-2500 br, 1761 cm⁻¹. ¹H NMR (300 MHz, CD₃SOCD₃) δ 5.85 (1H, s), 5.09 (1H, d, J = 4.6 Hz), 4.85 (1H, d, J = 5.1 Hz), 4.25-4.12 (2H, m), 2.99-2.82 (5H, m), 2.70 (1H, d, J = 5.1 Hz)18.2 Hz), 1.88-1.70 (3H, m), 1.63-1.50 (1H, m). ¹³ C NMR (75 MHz, CD₃OD) δ 176.21, 173.20, 114.16, 88.44, 65.54, 64.87, 62.71, 57.66, 52.27, 27.96, 27.89, 26.59. CIMS m/e 238 (MH+, base), 220. HRMS calcd. for C₁₂H₁₅NO₄ 237.1001. Found 237.1013. Anal calcd.for C₁₂H₁₅NO₄. C, 60.75; H, 6.37; N, 5.90. Found C, 60.73; H, 6.35; N, 5.68%. Further characterized as its diacetate and cyclic carbonate respectively. M.p. 143-145 (dec.). IR (KBr) 1762, 1737, 1653 cm⁻¹. ¹H NMR (300 MHz, CDCl₃) δ 1.7-2.1 (4H, m), 1.96 (3H, s, OCH₃), 1.99 (3H, s, OCH₃), 2.9-3.0 (3H, m), 3.1-3.2 (3H, m), 5.44 (1H, dd, J = 3.0, 8.5 Hz), 5.55 (1H, d, J = 8.5 Hz), 5.83 (1H, t, J = 1.7 Hz). CIMS, m/e 322 (MH+, 44), 321 (M+, 10), 280 (2), 262 (100), 221 (3), 202 (4), 177 (4), 154 (7). HRMS, m/e calcd. for $C_{16}H_{19}NO_6$ 321.1212, found 321.1205. Analysis, calcd. for C₁₆H₁₉NO₆, C, 59.81, H, 5.96, N, 4.36. Found C, 59.73, H 5.90, N, 4.09%.

M.p. decomposed between 170 and 235 °C. IR (KBr) 1817, 1771, 1658 cm⁻¹. ¹H NMR (300 MHz, CDCl₃) δ 1.8-2.0 (3H, m), 2.0-2.1 (1H, m), 2.8-3.0 (1H, m), 3.04 (2H, d, J = 2.4 Hz), 3.1-3.3 (2H, m), 3.53 (1H, s), 5.17 (2H, s), 5.94 (1H, t, J = 2 Hz). ¹³C NMR (75 MHz, CDCl₃) δ 171.7, 165.5, 153.3, 115.5, 83.8, 72.0, 70.7, 62.0, 51.9, 51.4, 27.3, 27.1, 25.7.

CIMS, m/e 264 (MH+, 100), 248 (12), 220, (18), 177 (4). HRMS, m/e calcd. for $C_{13}H_{13}NO_5$ 263.0794, found 263.0769. Analysis, calcd. for $C_{13}H_{13}NO_5$. C, 59.31, H, 4.98, N, 5.32. Found C, 59.02, H, 4.91, N, 5.00%.

Aldehyde 15. The alcohol 10 was converted into aldehyde acetonide 15 by catalytic osmylation (95%) (as above), acetonide formation (acetone/2,2-dimethoxy propane/HClO4, 90%) and oxidation (DMSO/oxalylchloride/Et₃N, 89%) to give 15, M.p 147°-149°C (from EtOAc). IR (CHCl₃) 2997, 1778, 1707, 1447, 1386, 1070 and 909 cm⁻¹. ¹H NMR (300 MHz, CDCl₃) δ 9.73 (1H, s), 5.84 (1H, s), 4.79 (1H, d, J = 7.6 Hz), 4.63-4.39 (1H, bs), 4.52 (1H, dd, J's = 7.6 and 1.5 Hz), 3.72 (3H, s), 3.61-3.50 (1H, bs), 3.10-2.92 (1H, dd, J's = 8.9 and 1.5 Hz), 2.89-2.66 (3H, m), 2.30-2.14 (1H, bs), 2.10-1.90 (1H, bs), 1.39 (3H, s), 1.35 (3H, s). ¹³ C NMR (75 MHz, CDCl₃) δ 200.71 (s), 171.92 (s), 167.50 (s), 155.94 (s), 114.58 (d), 111.30 (s), 85.53 (s), 72.57 (d), 71.36 (d), 58.65 (d), 53.29 (q), 49.63 (d), 41.17 (t), 26.59 (t), 25.59 (q), 24.26 (q), 24.06 (t). Anal calcd.for C₁₇H₂₁NO₇. C, 58.11; H, 6.03; N, 3.99. Found C, 57.94; H, 5.96; N, 3.90%. Crystals suitable for X-ray crystallographic analysis were grown from ethyl acetate.

Conversion of 16 into the alcohol 21. To a stirred solution of the diol 16 (250 mg, 1.05 mmol), DMAP (141 mg, 1.15 mmol) and Et_3N (0.16 ml, 1.15 mmol) in CH_2Cl_2 (20 ml) under argon was added trimethylacetylchloride (0.14 ml,1.10 mmol) dropwise. After 10 min the solvent was evaporated and the residues purified by chromatography on silica gel ($CH_2Cl_2/MeOH$, 10:1) to give a mixture of monoesters 17 and 18 (ratio 2:1) as a white solid (337 mg, 100%). Rfs = 0.38 and 0.45 (silica gel, 10:1 $CH_2Cl_2/MeOH$).

Oxalyl chloride (10.28 ml of 2M solution in CH_2Cl_2 , 20.56 mmol) and molecular sieves (4 Å) in dry CH_2Cl_2 (20 ml) were cooled to -78 °C under argon. DMSO (2.91 ml, 41.12 mmol) was then added dropwise so as to maintain an internal temperature below -60 °C. After 10 min a solution of the alcohols 17 and 18 (1.32 g, 4.11 mmol) in CH_2Cl_2 (12 ml) was added slowly. After 45 min, dry Et_3N (11.44 ml, 82.24 mmol) was added and the mixture stirred for a further 45 min. The cooling bath was removed and water added at room temperature. Stirring was continued for 10 min and the organic layer separated. The aqueous layer was extracted twice with CH_2Cl_2 (2x10 ml) and the combined organic layers dried (MgSO₄) before being concentrated in vacuo.

The crude ketones 19 and 20 from the above sequence were then dissolved in methanol (35 ml), cooled to 0 °C, and NaBH₄ (468 mg, 12.33 mmol) was added in small portions. The mixture was stirred for 15 min. Saturated NH₄Cl was added before neutralizing with 10% HCl. The methanol was evaporated in vacuo, and the aqueous phase extracted with CH₂Cl₂ (3x10 ml). The combined organic layers were dried (MgSO₄) before being concentrated. The crude product was purified by chromatography on silica gel (EtOAc) to give the desired product 21 as a white solid

(847 mg, 64% overall from 16). Rf = 0.43 (silica gel, EtOAc). M.p. 164-165 °C (dec.) (EtOAc/hexanes). IR (KBr): 3300-3100, 1773, 1725, 1663 cm⁻¹. ¹H NMR (300 MHz, CDCl₃) δ 1.16 (9H, s), 1.6-1.8 (1H, m), 1.85-2.10 (2H, m), 2.60-2.75 (1H, m), 2.83 (1H, dt, J = 19.2, 2.1 Hz), 2.95-3.20 (4H, m), 3.25-3.28 (1H, m), 3.45 (1H, br s), 3.63 (1H, br), 5.06 (1H, br s), 5.78 (1H, t, J = 1.8 Hz). ¹³C NMR (75 MHz, CDCl₃) δ 178.7, 173.2, 172.0, 112.0, 86.2, 76.3, 73.8, 61.5, 53.1, 51.4, 38.6, 28.4, 28.0, 27.0, 25.7. CIMS, m/e 322 (MH+, 100), 321 (M+, 28), 304 (5), 248 (5), 220 (83), 177 (9), 149 (2). HRMS, m/e calcd for C₁₇H₂₃NO₅ 322.1654. Found 322.1654.

Pentafluorophenylthiono ester 22. To a stirred solution of the alcohol 21 (150 mg, 0.467 mmol), DMAP (125 mg, 1.028 mmol) and molecular sieves (4 Å) in CH₂Cl₂ (5 ml) under argon was added pentafluorophenyl chlorothionoformate (0.15 ml, 0.935 mmol). After 30 min the solvent was evaporated and the residues purified by chromatography on silica gel (EtOAc/hexanes, 1:1, then 4:1) to give the thionoester 22 as a pale yellow oil (256 mg, 100%). Rf = 0.47 (silica gel, 4:1 EtOAc/hexanes). ¹H NMR (300 MHz, CDCl₃) δ 1.15 (9H, s), 1.7-1.9 (1H, m), 2.00-2.15 (2H, m), 2.35-2.47 (1H, m), 2.95 (1H, dt, J = 19.5, 2.1 Hz), 3.04-3.36 (5H, m), 5.42-5.46 (2H, m), 5.93 (1H, t, J = 1.8 Hz). ¹³C NMR (75 MHz, CDCl₃) δ 191.1, 176.8, 172.1, 169.4, 113.5, 86.3, 84.5, 69.8, 61.2, 53.4, 51.5, 38.5, 28.5, 27.8, 26.8, 25.5. CIMS, m/e 548 (MH+, 11), 446 (48), 364 (10), 322 (5), 304 (54), 276 (16), 220 (5), 213 (24), 185 (100), 184 (85), 147 (5), 129 (9), 117 (9), 102 (17). HRMS, m/e calcd for C₂₄H₂₃NO₆F₅S 548.1166. Found 548.1168.

Pivaloyloxynirurine 23. A stirred solution of the thionoester **22** (175 mg, 0.32 mmol), n-Bu₃SnH (0.26 ml, 0.96 mmol) and AIBN (8 mg) in benzene (8 ml) under argon was heated to reflux temperature for 15 min. The solvent was evaporated in vacuo and the residues purified by chromatography over silica gel (EtOAc-hexanes 1:1, then EtOAc) to give the desired product as a colorless oil (97 mg, 100%). Rf = 0.26 (silica gel, 4:1 EtOAc/hexanes). IR (CHCl₃) 1760, 1732, 1651 cm⁻¹. ¹H NMR (300 MHz, CDCl₃) δ 1.13 (9H, s), 1.25-1.40 (1H, m), 1.7-2.0 (4H, m), 2.81-3.13 (6H, m), 3.23 (1H, dd, J = 6.0, 2.7 Hz), 5.19-5.25 (1H, m), 5.73 (1H, t, J = 1.8 Hz). ¹³C NMR (75 MHz, CDCl₃) δ 177.4, 173.6, 173.2, 110.9, 83.5, 63.7, 62.1, 52.8, 51.2, 38.5, 31.5, 27.8, 27.1, 26.9, 25.5. CIMS, m/e 306 (MH+, 100), 305 (M+, 46), 291 (13), 269 (8), 235 (5), 233 (16), 213 (5), 204 (34), 184 (24), 177 (6), 135 (6), 111 (5), 103 (14). HRMS, m/e calcd for C₁₇H₂₃NO₄ 305.1627. Found 305.1628.

Hydroxynirurine 24. A solution of the ester **23** (92 mg, 0.287 mmol) and NaOMe (23 mg, 0.43 mmol) in methanol (3 ml) was stirred under argon at room temperature. After 5 h the solvent was evaporated and the residues purified by chromatography over silica gel (CH₂Cl₂:MeOH, 10:1) to give the corresponding alcohol **24** as a white solid (59 mg, 94%). Rf = 0.30 (silica gel, 10:1 CH₂Cl₂/MeOH). M.p. 153-155 °C (dec.). IR (KBr) 3097, 1758, 1650 cm⁻¹. ¹H NMR (300 MHz, CDCl₃) δ 1.32 (1H, m), 1.7-2.0 (4H, m), 2.44 (1H, br s), 2.71-3.10 (6H, m), 3.23 (1H, dt, J = 19.2, 2.1 Hz), 4.37-4.42 (1H, m), 5.67 (1H, t, J = 1.8 Hz). ¹³C NMR (75 MHz, CDCl₃) δ 175.4, 174.0, 110.3, 84.3, 62.0, 61.5, 55.7, 51.2, 34.1, 27.3, 27.2, 25.6. CIMS, m/e 222 (MH+, 100), 192 (25), 177 (6), 160 (14), 149 (34), 126 (8), 120 (35), 112 (53), 109 (6).

HRMS, m/e calcd for $C_{12}H_{15}NO_3$ 221.1052. Found 221.1055.

Norsecurinine 2. To a stirred solution of the alcohol 24 (5 mg, 0.023 mmol), Et₃N (7 μ l, 0.05 mmol) and DMAP (6 mg, 0.05 mmol) in CH₂Cl₂ (1 ml) under argon was added MeSO₂Cl (3.5 μ l, 0.045 mmol). After 15 min the solvent was evaporated and the residues purified by chromatography over silica gel (CH₂Cl₂/MeOH, 10:1) to give 2 as a white solid (4.2 mg, 91%). IR (CHCl₃) 1749, 1634 cm⁻¹. ¹H NMR (300 MHz, CDCl₃) δ 1.71 (1H, d, J = 11 Hz), 1.7-1.9 (2H, m), 1.98 (2H, m), 2.5-2.6 (2H, m), 3.21 (1H, t, J = 7.5 Hz) 3.28-3.33 (1H, m), 3.63 (1H, t, J = 5.5 Hz), 5.65 (1H, s), 6.48 (1H, d, J = 9.0 Hz), 6.74 (1H, dd, J = 9.0, 6.6 Hz). ¹³C NMR (75 MHz, CDCl₃) δ 172.8, 168.2, 143.5, 120.6, 108.0, 91.6, 65.1, 59.8, 55.3, 35.8, 29.3, 26.8. EIMS, m/e 203 (M⁺, 5), 134 (12), 106 (25), 78 (52), 70 (100), 61 (21), 55 (3), 51 (10), 43 (22). HRMS, m/e calcd for C₁₂H₁₃NO₂ 203.0946. Found 203.0957. The spectra were compared with an authentic sample of norsecurinine, kindly supplied by Dr. Peter Jacobi, and they were identical.

- 8-Hydroxynorsecurinine 26. A stirred solution of the diol 16 (300 mg, 1.27 mmol) in dry dioxane (10 ml) and molecular sieves (4 Å) under argon, was treated with a solution of Ph₃P (498 mg, 1.90 mmol) in dry THF (6 ml), followed by a solution of DEAD (331 mg, 1.90 mmol) in THF (6 ml). The mixture was stirred for 15 min at room temperature, and then warmed to 70-75 °C and stirred overnight at that The reaction mixture was diluted with methanol and the molecular sieves were filtered. The solvent was evaporated in vacuo and the residue purified by chromatography over silica gel (diethyl ether/methanol, 2:1) to give 8-hydroxy norsecurinine 26 (240 mg, 87%) as a white solid. M.p. decomposed above 165 °C. IR (KBr): 3500-3400, 1740, 1636 cm⁻¹. ¹H NMR (300 MHz, d₄-methanol) δ 1.65-2.10 (4H, m), 2.71-2.80 (1H, m), 3.23 (2H, dt), 3.60 (1H, t, J = 5.5 Hz), 4.71 (1H, d, J = 4.8)Hz), 5.81 (1H, s), 6.60 (1H, dd, J = 6.3, 9.0 Hz), 6.73 (1H, d, J = 9.3 Hz). ¹³C NMR (75) MHz, d₆-DMSO) δ 172.6, 166.6, 140.7, 121.0, 108.5, 92.4, 70.8, 62.9, 61.7, 54.6, 29.2, 26.6. CIMS, m/e 220 (MH+, 100), 219 (M+, 13), 202 (M-17, 18), 179 (7), 151 (2). Analysis, calcd. for C₁₂H₁₃NO₃. C, 65.74, H, 5.98, N, 6.39. Found C, 65.59, H, 5.91, N, 6.09%.
- 8-Hydroxynorsecurinine-N-oxide 27. To a stirred suspension of 8-hydroxynorsecurinine 26 (219 mg, 1 mmol) in methanol (15 ml) m-chloroperoxy benzoic acid was added (259 mg, 1.5 mmol). Immediately, the mixture became homogeneous. After 5 min. at room temperature the solvent was evaporated in vacuo and the residue purified by chromatography over silica gel (MeOH/CH₂Cl₂ 2:1, then MeOH) to give the N-oxide 27 (235 mg, 100%) as a white solid. M.p. decomposed between 155 and 178 °C. IR (KBr) 3608, 3450-3100, 1758, 1640 cm⁻¹. ¹H NMR (300 MHz, d₄-methanol) δ 2.10-2.50 (4H, m), 3.68 (2H, m), 3.82 (1H, m), 4.62 (1H, dd, J = 6.3, 4.5 Hz), 4.82 (1H, d, J = 4.5 Hz), 6.04 (1H, s), 6.43 (1H, dd, J = 9.0, 6.3 Hz), 7.05 (1H, d, J = 9.0 Hz). ¹³C NMR (75 MHz, d₄-methanol) δ 173.4, 165.1, 135.0, 126.8, 113.2, 88.9, 84.7, 74.3, 74.2, 72.8, 29.7, 25.8. CIMS, m/e 236 (MH+, 100), 235 (M+, 7), 220 (21), 218 (M-17, 27), 202 (6), 179 (9), 171 (4), 151 (5), 133 (7).

Rearrangement of 8-Hydroxynorsecurinine-N-oxide 27 into 28. A stirred suspension of 8-hydroxynorsecurinine N-oxide 27 (235 mg, 1 mmol) in xylenes (50 ml) was heated at reflux until all the solid was dissolved (approx. 30 min). The solvent was evaporated in vacuo and the residue purified by chromatography over silica gel (CH₂Cl₂/MeOH, 10:1) to give the desired product 28 (233 mg, 99%) as a white solid. M.p. decomposed between 165-199 °C (melt at 198-199 °C). IR (KBr): 3265, 1751, 1636 cm⁻¹. ¹H NMR (300 MHz, CDCl₃) δ 1.70-2.20 (4H, m), 2.81-2.92 (1H, m), 3.13 (1H, t, J = 8.7 Hz), 3.30-3.38 (1H, m), 4.49 (1H, t, J = 4.5 Hz), 4.62 (1H, t), 5.91 (1H, s), 6.22 (1H, dd, J = 9.3, 5.7 Hz), 7.03 (1H, d, J = 9.3 Hz). ¹³C NMR (75 MHz, CDCl₃) δ 172.1, 163.9, 131.1, 127.1, 114.1, 85.5, 74.4, 72.4, 67.78, 57.0, 24.6, 23.5. CIMS, m/e 236 (MH+, 100), 235 (M+, 14), 218 (46), 190 (12), 179 (50), 172 (9), 151 (47), 133 (85). Analysis, calcd. for C₁₂H₁₃NO₄. C, 61.27, H, 5.57, N, 5.95. Found C, 61.12, H, 5.51, N, 5.70%.

Prenirurine 34. The procedure adopted for the Swern oxidation was similar to that described previously. The crude ketone 33 obtained was reduced with NaBH₄ at 0 °C for 15 min. After evaporating the solvent the crude product was directly purified by chromatography on silica gel (CH₂Cl₂/MeOH, 10:1) to give the desired product 34 as a white solid (82% overall). Rf = 0.18 (silica gel, 10:1 CH₂Cl₂/MeOH). M.p. 146-148 °C (dec). IR (KBr) 3129, 1767, 1654 cm⁻¹. ¹H NMR (300 MHz, CDCl₃) δ 1.65-1.80 (1H, m), 1.85-2.01 (2H, m), 2.04-2.22 (2H, m), 2.35 (1H, dd, J = 13.5, 4.5 Hz) 2.46 (1H, br s), 2.65 (1H, dt, J = 19.2, 2.1 Hz), 2.98 (1H, br s), 3.05-3.15 (2H, m), 3.32 (1H, s), 3.65-3.74 (1H, m), 4.22 (1H, dd, J = 10.5, 4.5 Hz), 5.74 (1H, s). ¹³C NMR (75 MHz, CDCl₃) δ 174.2, 173.7, 111.6, 84.6, 72.2, 63.2, 56.2, 52.7, 33.1, 31.2, 26.9, 25.6. CIMS, m/e 222 (MH+, 100), 221 (M+, 9), 206 (2), 185 (15), 171 (4), 157 (15), 153 (4), 129 (6), 127 (6), 125 (37), 111 (12). HRMS, m/e calcd for C₁₂H₁₅NO₃ 221.1052. Found 221.1054.

Oxidation of Prenirurine. To a solution of 34 (2.5 mg) in dichloromethane (1.0 ml) at 25 °C was added m-chloroperoxybenzoic acid (3 mg), and the mixture stirred for 1 h. The solution was treated with trifluroacetic anhydride (3 μ l) and stirring continued for 15 h. The solution was diluted with dichloromethane (3 ml), washed with saturated aqueous NaHCO₃, dried (Na₂SO₄), and evaporated to give a mixture (1.5 mg). The ¹H NMR of the mixture was complex, but characteristic signals at 5.94 (1H, s), 6.28 (1H, dd, J = 9.3, 5.7 Hz) and 6.60 (1H, d, J = 9.3 Hz) indicated the presence of 38 (major component), and a diagnostic signal at 4.96 (1H, d, J = 2.8 Hz, H_{2a}) for nirurine 1 (<10%).

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